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Approach to a Comparative Study of the Metabolism of Porphyrins and Chlorophylls

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Abstract

Metabolic pathways are series of successive biochemical reactions catalyzed by enzymes which together constitute a process of extraordinary complexity, the cellular metabolism. Like any other biological phenomenon, metabolism is the result of evolution. Different techniques are used to compare characteristics of living entities seeking to analyze the similarity between them. The features or characteristics that define species are of different types: molecular, metabolic, cellular, environmental, behavioral, etc. Depending on the similarity, biological species and any entity of the biological hierarchy, are sorted and classified. This paper focuses on studying a characteristic of organisms that is metabolism of porphyrins and chlorophylls using cladistic procedures, unlike others, allow crawl the changes in this feature. The result of a cladistic analysis is an evolutionary hypothesis, a hypothesis about the evolution of metabolism of porphyrins and chlorophylls. The work raises two objectives: firstly, hypotheses about the evolution of metabolism of porphyrins and chlorophylls; otherwise, hypotheses on the evolution of enzymes involved in cellular metabolism this part, opening the possibility that some of them serve as phylogenetic marker.

Keywords: porphyrins and chlorophylls metabolism, cladistics, biological evolution

1. Introduction: A look back in time

Metabolic pathways are a series of successive biochemical reactions catalyzed by enzymes which together constitute a process of extraordinary complexity, cellular metabolism. Like any other biological phenomenon, metabolism is the result of evolution.

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As the history of the species and their family relationships are derived from comparative studies, the history of metabolism can only be reconstructed through comparative analysis of its elements (Cunchillos and Lecointre, 2002, 2003, 2005).

Using the cladistic analysis to build an evolutionary hypothesis of the emergence and evolution of porphyrin and chlorophyll metabolism involves considering photosynthesis from the evolutionary point of view. On the origin of photosynthesis there are basically two hypotheses (Olson, 1999, 2001): one suggests that it originated in the prebiotic atmosphere of the Earth and comes to life; another, based on recent molecular phylogenetic analysis suggests that photosynthesis arose after chemolithotrophs organisms appear. However, we know little about how they were incorporated into the photosynthetic process its multiple biochemical components.

The scientific community agrees that photosynthesis is a process that originates in bacteria because, firstly, there is no archaea with Mg-tetrapyrroles based photosystems and, secondly, photosynthetic eukaryota acquired this ability from cyanobacteria through endosymbiosis. It is possible to achieve a better understanding of the evolution of photosynthesis comparatively studying genes or gene products of photosynthesis in the groups of bacteria that synthesize photosystems: purple bacteria (proteobacteria, with photosystem II), green non-sulfur bacteria (photosystem II), green sulfur bacteria (photosystems I and II). The Earth is about 4,500 million years and the beginning of life is in the Archean period, about 3800-4000 million years ago. Stromatolites (the oldest Archaean fossils) containing evidence of biological carbon fixation have an approximate age of 3,800 million years (Schidiowski, 1988) while the oldest cyanobacteria-like cell microfossils have an age of 3,500 million years (Schopf, 1993; Schopf and Packer 1987), ie, appearing 300 million years after the first record of life.

However, Earth was dominated at first by cyanobacteria which were 2,700 million years ago, coinciding with the first appearance of oxygen on earth (Des Marais, 2000). During approximately 1 million years, cyanobacteria increased the oxygen level to almost a quarter of the current level. The success of these organisms could be due not only to the power advantage of photosynthesis but due also to inhibition by oxygen (as toxicant) of potential competitors.

The emergence of the first eukaryotes occurred about 1,800 million years ago, a fact that required 600-800 million years for cyanobacteria were incorporated by endosymbiosis in eukaryotes and evolved into chloroplasts. Thereafter the algae increased the oxygen level until present. Finally, his descendants land plants appeared 500 million years ago.

There is general consensus on the hypothesis suggesting the evolution of photosynthetic pigments from chemoautotrophs: they lived in an environment of chemical imbalance that led to the ability to use the pigments to harness light as an additional source of energy. Established photosynthesis, this might evolve to allow cells use sunlight as the only energy source (Nisbert and Sleep, 2001). On this tour it must be some intermediate stage that could be starring by purple bacteria: the anoxygenic photosynthesis bacteria evolve from this group who had phototaxis by infrared light (Nisbert et al., 1995).

This hypothesis is based on the close relationship between the emission spectrum of geothermal light and absorption spectrum of bacteriochlorophylls b, relationship that serves the authors to raise the possibility that photosynthesis arose in organisms presenting bacteriochlorophylls a or b. They lived in the vicinity of oceanic hydrothermal vents where they could detect a faint infrared radiation.

Accordingly, these phototaxis bacteria with the ability to detect infrared light have lived in an optimum environment which would have been an evolutionary advantage in terms of competition for resources or nutrients. Subsequently adaptation of this primitive photosystem would have allowed organisms to start using the far-red sunlight as it moved into shallower waters. Over time, chlorophylls appear to use light of higher energy (visible) and water photolysis. Photosynthesis in bacteria and plants occurs in two phases, photochemistry, and biochemistry. Photochemical phase involves the participation of photosynthetic pigments including chlorophylls and bacteriochlorophylls of bacteria and plants which are derived from the porphyrin metabolism.

2. Map of metabolism of porphyrins and chlorophylls

Porphyrins are tetrapyrroles binds covalently to a metal: iron (Fe) to form cytochromes, peroxidase, catalase, myoglobin and hemoglobin; copper (Cu) or nickel (Ni) to form molecules for electron transport in methanogenic bacteria; magnesium (Mg) to form chlorophylls and bacteriochlorophylls.

In the porphyrin biosynthesis pathway (Fig. 1) there are two stages:

1) Synthesis of 5-aminolevulinate (ALA): In all photosynthetic eukaryota and prokaryota, except the α -proteobacteria, ALA is synthesized from glutamate which binds glutamyl-tRNA by glutamyl- tRNA synthase catalyzed reaction (GluRS). Moreover, all non-photosynthetic eukaryota (animals, fungi and apicomplexa) and α -proteobacteria form ALA by condensing glycine and succinyl-CoA in a reaction catalyzed by ALA synthase.

2) Synthesis of protoporphyrin IX from 5-aminolevulinate (ALA), a chain of six reactions, the same in all organisms

Figure 1: Porphyrins and chlrophylls biosynthesis pathway (Modified from GenomeNet) (red arrows indicate enzymes that undergo a single change in the analysis)



In the context of evolution it is important to highlight the conceptual difference between "evolution of photosynthesis" and "evolution of photosynthetic organisms": the evolution of photosynthesis involves a limited number of genes and / or gene products while the evolution of photosynthetic organisms involves the complete genome. The phylogeny of the 16S subunit rRNA (Woese, 1987) establishes three domains: Archaea (archaebacteria), Bacteria (eubacteria) and Eucarya (eukaryotes). Although this classification was discussed by other authors (Cavalier-Smith, 1992), the fact is that other phylogenetic studies using new genomic and sequence data provide consistent results with all three domains. However, when phylogenetics are compared, inconsistencies appear showing that the former do not necessarily reflect the second: evolution of organisms *versus* evolution of genes.

3. Cladistic analysis to approach the evolution of porphyrins metabolism

For the comparative study of various types of entities (molecules, metabolism, organs, organisms, populations, behaviors, geographic distribution, ecosystems) different techniques are used all aimed at analyzing the similarity between the strains under study. Between different procedures, cladistic analysis provides a high degree of objectivity to study but all techniques provide useful and necessary information in the evolutionary context.

The porphyrins and chlorophyll metabolism is a complex process involving numerous chemical reactions catalyzed by enzymes. Its structure (components and functions) in the diversity of organisms is a product of evolution and can only be reconstructed by comparing the components. Tetrapyrroles serve as electron carriers in all domains of life. Heme carrying proteins were postulated to have been present in the last common ancestor of Bacteria and Archaea.

The application of the comparative method requires first determine the set of elements to compare. These elements are referred to as cladistic terminology "operational taxonomic units". They can refer to molecules or species, taxa in general. In this particular case, the set of taxa to be compared is the metabolism of porphyrins and chlorophylls in a set of organisms that constitute the study group (Table 1). Taxon, that is, each organism metabolism study group, is defined by characters that are enzymes (Table 2) involved in this part of cellular metabolism (Fg.1).

Enzymes can be present or absent in a particular taxon (the bchl gene is absent in archaebacteria and present in bacteria) which means that the same character may have different "versions" which are called "character state".

ACRONYM	NAME OF THE SPECIES	DOMAIN	GROUP		
afu	Archaeoglobus fulgidus	Archaea	euryarchaea		
ago	Ashbya gossypii	Eukarya	fungi		
ana	Nostoc	Bacteria	cyanobacteria		
ape	Aeropyrum pernix	Archaea	crenarchaea		
ath	Arabidopsis thaliana	Eukarya	plant		
atu	Agrobacterium fabrum	Bacteria	α-proteobacteria		
bja	Bradyrhizobium diazoefficiens	Bacteria	α-proteobacteria		
bsu	Bacillus subtilis	Bacteria	firmicutes		
buc	Buchnera aphidicola	Bacteria	γ-proteobacteria		
cac	Clostridium acetobutylicum	Bacteria	firmicutes		
cal	Candida albicans	Eukarya	fungi		
cau	Chloroflexus aurantiacus	Bacteria	chloroflexi		
cch	Chlorobium chlorochromatii	Bacteria	chlorobi		
cel	Caenorhabditis elegans	Eukarya	metazoa		
cho	Cryptosporidium hominis	Eukarya	alveolata		
cme	Cyanidioschyzon merolae	Eukarya	rodofite		
cne	Cryptococcus neoformans	Eukarya	fungi		
cte	Chlorobium tepidum	Bacteria	chlorobi		
ddi	Dictyostelium discoideum	Eukarya	amebozoa		
det	Dehalococcoides mccartyi	Bacteria	chloroflexi		
dge	Deinococcus geothermalis	Bacteria	deinococus		
dme	Drosophila melanogaster	Eukarya	metazoa		
eco	Escherichia coli	Bacteria	γ-proteobacteria		
gox	Gluconobacter oxydans	Bacteria	α-proteobacteria		
gvi	Gloeobacter violaceus	Bacteria	cyanobacteria		
hal	Halobacterium sp. NRC-1	Archaea	euryarchaea		
hbu	Hyperthermus butylicus	Archaea	crenarcheae		
hin	Haemophilus influenzae	Bacteria	γ-proteobacteria		
lic	Leptospira interrogans	Bacteria	spirochetae		
lpf	Legionella pneumophila	Bacteria	γ-proteobacteria		
mac	Methanosarcina acetivorans	Archaea	euryarchaea		
mca	Methylococcus capsulatus	Bacteria	γ-proteobacteria		
mga	Mycoplasma gallisepticum	Bacteria	mollicutes		
mja	Methanocaldococcus jannaschii	Archaea	euryarchaea		
mka	Methanopyrus kandleri	Archaea	euryarchaea		
mth	Methanothermobacter	Archaea	euryarchaea		
	thermautotrophicus				
mtu	Mycobacterium tuberculosis	Bacteria	actinobacteria		
neq	Nanoarchaeum equitans	Archaea	nonarchaea		
neu	Nitrosomonas europaea	Bacteria	β-proteobacteria		
nwi	Nitrobacter winogradskyi	Bacteria	α-proteobacteria		
pab	Pyrococcus abyssi	Archaea	euryarchaea		
pac	Propionibacterium acnes	Bacteria	actinobacteria		

Table 1: Study Group, metabolism of porphyrins and chlorophylls in 63 species

pae	Pseudomonas aeruginosa	Bacteria	γ-proteobacteria
pai	Pyrobaculum aerophilum	Archaea	crenarchaea
pfa	Plasmodium falciparum	Eukarya	alveolata
plt	Prevotella intermedia	Bacteria	bacteroidete
ppr	Photobacterium profundum	Bacteria	γ-proteobacteria
rme	Cupriavidus metallidurans	Bacteria	β-proteobacteria
rpa	Rhodopseudomonas palustris	Bacteria	α-proteobacteria
rpr	Rickettsia prowazekii	Bacteria	α-proteobacteria
rru	Rhodospirillum rubrum	Bacteria	α-proteobacteria
sce	Saccharomyces cerevisiae	Eukarya	fungi
SSO	Sulfolobus solfataricus	Archaea	crenarchaea
syc	Synechococcus elongatus	Bacteria	cyanobacteria
syn	Synechocystis sp.	Bacteria	cyanobacteria
tac	Thermoplasma acidophilum	Archaea	euryarchaea
tbd	Thiobacillus denitrificans	Bacteria	β-proteobacteria
tcr	Trypanosoma cruzi	Eukarya	euglenozoa
tde	Treponema denticola	Bacteria	spirochetae
tel	Thermosynechococcus elongatus	Bacteria	cyanobacteria
tth	Thermus thermophilus	Bacteria	deinococcus
zmo	Zymomonas mobilis	Bacteria	α-proteobacteria

Table 2: Enzymes, characters selected in the study group

Data code	ENZYME NAME	EC	GENE NAME
B/ 1	5-aminolevulinate synthase	2.3.1.37	ALAS, HemA
C/ 2	glutamyI-tRNA synthetase	6.1.1.17	gltX, others
D/ 3	glutamyl-tRNA reductase	1.2.1.70	HemA
E/4	glutamate-1-semialdehyde 2,1-aminomutase	5.4.3.8	HemL
F/5	porphobilinogen synthase	4.2.1.24	HemB
G/ 6	hydroxymethylbilane synthase	2.5.1.61	HemC
H/ 7	uroporphyrinogen-III synthase	4.2.1.75	HemD
1/8	uroporphyrinogen decarboxylase	4.1.1.37	HemE
J/ 9	uroporphyrinogen methyltransferase	2.1.1.107	HemX, CysG, CobA
K/ 10	precorrin-2 dehydrogenase	1.3.1.76	CysG
L/ 11	sirohydrochlorin ferrochelatase	4.99.1.4	CysG
M/ 12	sirohydrochlorin cobaltochelatase	4.99.1.3	CbiX, CbiK
N/ 13	cobalt-factor II C20-methyltransferase	2.1.1.151	CbiL, CbiL, CbiF
O/ 14	precorrin-3 methyltransferase	2.1.1.131	CbiH, CbiL,CobJ
P/ 15	precorrin-3 methylase	2.1.1.133	CbiF, CobM
Q/ 16	precorrin-6A reductase	1.3.1.54	CobL, CobK
S/ 17	precorrin-6 methyltransferase	2.1.1.132	CbiE, CbiT, CobL
T/ 18	precorrin isomerase	5.4.1.2	CobH, CbiC
U/ 19	cob(II)yrinic acid a,c-diamide reductase	1.16.8.1	
V/20	cob(I)yrinic acid a,c-diamide	2.5.1.17	CobO, CobA, BtuR

r			
14/ 04	adenosyltransterase		
W/ 21	adenosylcobyric acid synthase (glutamine-	6.3.5.10	CDIP, CODQ
<u>)// 00</u>	nyaroiysing	(0.1.10	
X/22	adenosylcobinamide-phosphate synthase	6.3.1.10	CobD, CbiB
Y/ 23	Adenosylcobinamide kinase	2.7.1.156	CobU, CobP
Z/ 24	adenosylcobinamide-GDP	2.7.8.26	CobS
	ribazoletransferase		
AA/ 25	alpha-ribazole phosphatase	3.1.3.73	CobC
AB/ 26	cob(I)yrinic acid a,c-diamide	2.5.1.17	
	adenosyltransferase		
AC/ 27	aquacobalamin reductase	1.16.1.3	
AD/ 28	precorrin-2 C20-methyltransferase	2.1.1.130	CbiL, Cobl
AE/ 29	precorrin-3B synthase	1.14.13.83	CobG
AF/ 30	precorrin-6A synthase (deacetylating)	2.1.1.152	CobF
AG/ 31	coproporphyrinogen oxidase	1.3.3.3	HemF
AH/ 32	coproporphyrinogen dehydrogenase	1.3.99.22	HemN
AJ/ 33	protoporphyrinogen oxidase	1.3.3.4	HemG
AK/ 34	protoporphyrin IX magnesium-chelatase	6.6.1.1	Chld, Bchl
AL/ 35	magnesium protoporphyrin IX	2.1.1.11	Chl, Bchl
	methyltransferase		
AM/ 36	Mg-protoporphyrin IX monomethyl ester	1.14.13.81	Acs, Pni
	(oxidative) cyclase		
AN/ 37	protochlorophyllide reductase	1.3.1.33	
AO / 38	chlorophyllase	3.1.1.14	
AP / 39	chlorophyll synthase	2.5.1.62	
AQ / 40	ferrochelatase	4.99.1.1	HemH
AR/ 41	ferroxidase; ceruloplasmin	1.16.3.1	
AS / 42	cytochrome c heme-lyase	4.4.1.17	
AT / 43	heme oxygenase	1.14.99.3	
AU / 44	phytochromobilin:ferredoxin	1.3.7.4	
	oxidoreductase		
AV / 45	phycocyanobilin:ferredoxin oxidoreductase	1.3.7.5	
AW / 46	15.16-dihydrobiliverdin:ferredoxin	1.3.7.2	
	oxidoreductase		
AX / 47	phycoerythrobilin ferredoxin	1373	
	oxidoreductase		
AY / 48	biliverdin reductase	1.3.1.24	
A7 / 49	bilirubin oxidase	1.3.3.5	
BA / 50	bilirubin UDP-ducuronosyltransferase	24117	
BB / 51	beta-alucuronidase	32131	
557 51	sola giudai di lidaso	0.2.1.01	

The absence or presences of an enzyme in the metabolism of organisms that constitute the study group are discrete characters that are encoded in a binary system, 0 or 1. This coding means that an enzyme can be in two states: 0-absent or 1-present.

Data have been collected from KEGG: Kyoto Encyclopedia of Genes and Genomes (1995-2015 Kanehisa Laboratories)

The set of taxa and characters are reflected in a "matrix data" showing taxa in rows and states of a character in columns (Tale 3). This matrix is the basis for further analysis.

Table 3: Data Matrix showing the states of the characters (0-absent enzyme, 1-present enzyme)



Table 3. cont.

1 1	S	Т	U	V	W	х	Y	Z	AA	AB	AC	AD	AE	AF	AG	AH
	2.1.1.132	5.4.1.2	1.16.8.1	2.5.1.17	6.3.5.10	6.3.1.10	2.7.1.156	2.7.8.26	3.1.3.73	2.5.1.17	1.16.1.3	2.1.1.130	1.14.13.83	2.1.1.152	1.3.3.3.	1.3.99.22
ath	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
cel	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
cal	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
cho	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
cme	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
dme	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
pfa	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
sce	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
ape	Ő	0	Ő	1	0	1	Ő	1	0	1	Ö	Ő	Ő	0	Ő	0
afu	1	1	0	0	1	1	0	1	0	0	0	0	0	0	0	0
hbu	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
mth	1	1	0	0	1	1	0	1	0	0	0	0	0	0	0	0
mja mka	1	1	0	0	1	1	0	1	0	0	0	0	0	0	0	0
mac	1	1	0	1	1	1	0	1	0	1	0	0	0	0	0	0
neq	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
pab	0	0	0	1	0	1	0	1	0	1	0	0	0	0	0	0
\$90	1	1	1	0	1	1	0	1	0	0	0	1	0	0	0	0
ana	1	1	0	1	1	1	1	1	0	1	0	1	0	0	1	1
gvi	1	1	0	1	1	1	1	1	0	1	0	1	0	0	1	1
syc	1	1	0	1	1	1	1	1	0	1	0	1	0	0	1	1
tel	1	1	0	1	1	1	1	1	0	1	Ö	1	0	Ö	1	1
atu	1	1	1	1	1	1	1	1	0	1	0	1	1	0	1	1
gox	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
nwi	0	0	0	1	0	0	0	0	0	1	0	0	0	0	1	1
rpa	1	1	1	1	1	1	1	1	0	1	0	1	0	1	1	1
rpr	0	Ó	0	0	0	Ö	Ö	0	Ő	Ö	Ő	0	Ő	0	1 1	1
zmo	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
rme	0	0	1	1	1	1	1	1	Ő	1	0	0	Ő	0	1	1
tbd	0	0	1	1	1	1	1	1	0	1	0	0	0	0	1	1
eco	0	0	0	1	0	0	1	1	1	1	1	0	0	0	1	1
hin	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
Ipf	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
ppr	0	0	Ö	1	1	1	1	1	0	1	1	0	0	0	1	1
pae	1	1	1	1	1	1	1	1	0	1	0	1	1	0	1	1
cac	1	1	0	0	1	1	1	1	1	0	0	0	0	0	0	1
mga	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
tth	1	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0
dge	1	1	0	1	1	1	1	1	Ő	1	Ő	1	Ő	Ő	Ő	1
det	1	0	0	1	1	1	1	1	1	1	0	0	0	0	0	1
cch	0	0	1	1	1	1	1	1	0	1	0	0	0	0	0	1
plt	0	0	1	1	1	1	1	1	0	1	0	0	0	0	0	1
pac	0	1	0	1	1	1	1	1	0	1	0	1	0	1	0	1
lic	1	1	0	1	1	1	1	1	0	1	0	1	0	0	1	0
cau	1	0	0	1	0	0	1	0	0	1	0	0	0	0	0	1
AI	AJ	AK A	AL AM	AN	AO	AP	AQ	AR A	S AT	AU	AV	AW	AX	AY A	AZ BA	BB
ath	1.3.3.4	6.6.1.1 2.1	1.11 1.14.13	81 1.3.1.33	3.1.1.14	2.5.1.62	4.99.1.1 1.	16.3.1 4.4.1	1.17 1.14.99.	3 1.3.7.4	1.3.7.5	1.3.7.2	1.3.7.3 1.3	0 1.3	3.3.5 2.4.1.1	3.2.1.31
ago	1	0	1 0	0	0	0	1	0 1	0	0	0	0	0	0	0 1	0
cal	1	0	1 0	0	0	0	0	0 0	0	0	0	0	0	0	0 1	0
cne	1	0	0 0	0	0	0	1	0 1	0	0	0	0	0	0	0 1	0
cme	1	1	1 0	1	0	1	1	0 0	1	0	0	0	0	0	0 0	0
ddi	1	0	1 0	0	0	0	1	0 1	0	0						0
pfa	1	0	1 0	0	0	0	1	0 1	1	0	0	0	0	1	0 1	1
tor	0	0	1 0	0	0	0	1	0 1	1	0	0	0	0	0	0 1 0 1 0 0	1
ape	0	0	1 0 0 0 1 0 0 0	0	0 0 0 0 0	0 0 0 0 0 0	1 1 1 0	0 1 0 1 0 1 0 0	1 0 0	0 0 0 0 0	0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	0 0 0 0	0	0 0 0 0 0 0	0 1 0 1 0 0 0 1 0 0	0 0 0
hal	0	0 0 0 1	1 0 0 0 1 0 0 0 0 0 0 0	0	0	0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	1 1 0 0		1 0 0 0	0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0		0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0		0 1 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	0 1 0 1 0 0 0 1 0 0 0 0 0 0	0
	0	0 0 0 1 1	1 0 0 0 1 0 0 0 0 0 0 0 1 0	0 0 0 0 0 0			1 1 0 0 0 0 0		1 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	0 0 0 0 0 0 0	0 0 0 0 0 0 0	0 1 0 0 0 0 0 0 0 0 0 0 0 0 0	0 1 0 0 0 1 0 0 0 0 0 0 0 0 0 0 0 0	0 0 0 0 0 0 0
mth	0	0 0 1 1 0 1	1 0 0 0 1 0 0 0 0 0 0 0 1 0 0 0 0 0 0 0		0 0 0 0 0 0 0	0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	1 1 0 0 0 0 0 0			0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0				0 1 0 0 0 0 0 0 0 0 0 0 0 0 0	0 1 0 1 0 0 0 1 0 0 0 0 0 0 0 0 0 0 0 0	
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A data matrix as the above in which the reflected characters are metabolic enzymes in two states, present (1) or absent (0), serves for the cladistic analysis of enzymes and accordingly the metabolism of porphyrins and chlorophylls in organisms of the study group.

The cladistic analysis was performed with the MIX program in Phylip (Felsestein, 2001) (<u>http://evolution.genetics.washington.edu/phylip.html</u>) which estimates phylogenies by Wagner parsimony method for discrete character data with two states (0 and 1) (also available an online version of the program <u>http://mobyle.pasteur.fr/cgi-bin/portal.py#jobs::overview</u>). Cladograms and trees are drawn with PHY.FI application (Fredslund, 2006).

The result of this analysis is reflected by a cladogram representing an evolutionary hypothesis discussed below.

4. One hypothesis about the evolution of porphyrins and chlorophylls

The cladistic analysis following Wagner Parsimony Method for discrete characters in two states provides 100 equally parsimonious trees (L = 233). Figure 2 shows the majority rule consensus tree and Figure 3 shows the strict consensus tree of the 100 most parsimonious and equally parsimonious trees.

In the majority rule consensus tree two large groups or clades are observed: one formed by Bacteria and Archaea and another group of Bacteria and Eukaryota (Fig. 2). This result is confirmed by the strict consensus tree (Fig. 3). The ancestral metabolism corresponds to *Nanoarchaeum equitans* (neq) leaving immediately after the ancestor (anc, the hypothetical ancestor with all characters set to 0, plesiomorphic, ie, absent).

All enzymes considered undergo at least one change of state. Those experiencing a single change (bold in the Table 2) are apomorphic characters which define well metabolism (red arrows in Fig. 1). All other enzymes experience more than one change, they are homoplasy, indicate convergent evolution, the adaptive nature of metabolism and its components.

In the clade Metabolism **[BACTERIA + ARCHAEA]** the group of cyanobacteria (tel, gvi, syn, ana and syc) is well defined by the enzyme **phycocyanobilin:ferredoxin oxidoreductase (EC 1.3.7.5)** which experience a unique change and is a sinapomorphic character for the cyanobacteria clade. Within this group of cyanobacteria, *Gloeobacter violaceus* (gvi) is defined by two apomorphic characters: enzymes **15,16-dihydrobiliverdin:ferredoxin oxidoreductase (EC 1.3.7.3)**.

On the basis of this group of cyanobacteria, in a previous node, is the metabolism of *Chloroflexus aurantiacus* (cau) defined by an autoapomorphy, the **bilirubin oxidase** enzyme (EC 1.3.3.5). The clade Metabolism **[BACTERIA + EUKARYA]** includes among eukaryotes *Arabidopsis thaliana* (ath), plant whose metabolism is well defined by two apomorphies, enzymes **chlorophyllase (EC 3.1.1.14)** and **phytochromobilin: ferredoxin oxidoreductase (EC 1.3.7.4)**.

All other groups or clades are not based on apomorphic characters, no homologies to well define species metabolism. Therefore they are analogies and a case of convergent or parallel evolution. The similarity is due to homology and analogy. But nevertheless analogies or homoplasies not serve to discover phylogenetic relationships. Therefore they can not be established assumptions about relationships between sister groups based on the metabolism of porphyrins and chlorophylls.

Resulting cladograms both majority consensus as strict consensus (Figs. 2 and 3) reflects relationships derived from adaptive changes that lead to the expression, or not, of a gene and possible synthesis or catalytic activity of an enzyme as well as the acquisition or loss metabolic capabilities as an adaptive response to the environment. Moreover, lateral gene transfer is a key process in the early stages of cellular evolution that draws a complex network of relationships that mask and / or replace the vertical transfer and evolution model based on ancestry with modification. On the other hand, much of the diversity found in algae is due to secondary and tertiary endosymbiotic events (Keeling, 2010).



Figure 2. Mayority rule consense tree

Figure 3. Strict consense tree

The metabolism compared in the study group makes clear enzymes that change once. Some of these elements as apomorphic homologies could be phylogenetic markers. This is the case of phycocyanobilin:ferredoxin oxidoreductase (EC 1.3.7.5) for cyanobacteria and chlorophyllase (EC 3.1.1.14) for plants.

Green algae and its progeny lost the cyanobacterial phycobilisome lightharvesting system, whereas red algae (cme, *Cyanidioschyzon merolae*) (on the cladogram with *Arabidopsis thaliana*) and glaucophytes retained it. The chloroplasts in algae and plants are derived from cyanobacteria and endosymbiosis gave rise to photosynthetic eukaryotes (Margulis, 1992). The evolution of photosynthesis is a complex process involving different sources and routes of its many components, so that its history can not be described as a simple, linear process. However, it seems certain that the emergence of Mg-tetrapyrrole and apoproteins of the reaction centers are key events that led to the development of the photosynthetic process. The flow of electrons acceptor–is channeled by protein complexes that always contain metallo-organic cofactors. Membrane-bound complexes couple the transfer of electrons across the membrane to the generation of an ion gradient and transmembrane electrical potential. This chemiosmotic mechanism was likely present in the last common ancestor and has been carried forward to the three presently persisting domains of life, Bacteria, Archaea, and Eukarya (Lane et al., 2010).

The Granick (Granick, 1965) and Retrograde (Horowitz, 1945) hypotheses on the establishment of metabolic pathways are complementary. Granick established as hypotheses that the intermediate compounds of the modern biosynthetic pathways were the final products of early pathways and thus the evolution of the pathway can be traced from the beginning to the end. However this contrasts with the retrograde hypothesis which posits that present biosynthetic pathways are set up in the reverse order to their evolutionary history and occurred through gene duplications. But The Granick and retrograde hypotheses are not mutually exclusive because the retrograde hypothesis is a consequence of the depletion of base molecules present in the primordial soup, molecules that follow the Granick hypothesis may be more derived.

There are still many aspects of the evolution of photosynthesis unresolved due in part to the existence of highly diversified components (Fig.4). One way to learn more about this issue is to address the systematic, descriptive and comparative study of genes and gene products of photosynthesis in the diversity of phototrophic organisms. For this purpose contributes undoubtedly the availability of numerous molecular data and the use of phylogenetic analysis tools. For example and based on the results of this study, comparative analysis of enzymes glutamyl-tRNA synthetase (EC 6.1.1.17), porphobilinogen synthase (EC 4.2.1.24) and protoporphyrinogen oxidase (EC 1.3.3.4) arises With all this new knowledge will continue to emerge thus reconstruct the evolutionary history of photosynthesis.



Figure 4: Some molecules of metabolism of porphyrins and chlorophylls

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